

Figure 1. Schematic model for the binding of an acyl pyrophosphate as a substrate analogue.

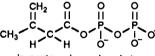
Amines and thiols are acylated by acyl pyrophosphates in organic solvents and in aqueous solution.

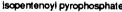
$$CH_{3} \xrightarrow{C} O \xrightarrow{P} O \xrightarrow{P} O \xrightarrow{P} O \xrightarrow{R} + R^{NH_{2}} \xrightarrow{O} CH_{3} \xrightarrow{O} \xrightarrow{N} \xrightarrow{R} + PP_{i}$$

Reaction of acyl pyrophosphates (50 mM) with carboxyl-terminal-protected amino acid esters (250 mM) in phosphate buffer (200 mM) at pH 7.0 produces N-acylated amino acid esters (63% yield after workup and purification of reaction of valine ethyl ester and phenylacetyl pyrophosphate). Lysine ethyl ester is preferentially acylated at the ϵ -amino group under these conditions.

Acyl pyrophosphates hydrolyze in wet organic solvents or in water to form inorganic pyrophosphate and the corresponding acids. Inorganic pyrophosphate does not incorporate ¹⁸O from the hydrolysis of 0.25 M isopentenoyl diphosphate in 0.75 M pH 7.0 HEPES buffer at 37 °C containing 25% H₂¹⁸O (³¹P NMR ¹⁸O analysis^{16–18}). Thus, the hydrolysis occurs by acylation of water with pyrophosphate as the intact leaving group. Between pH 3.0 and 7.0, the hydrolysis of phenylacetyl pyrophosphate follows first-order kinetics (monitored by ³¹P NMR), $k_{obsd} = 2.7$ \pm 0.5 × 10⁻⁵ s⁻¹, pH 6.2, 37 °C, 0.5 M potassium maleate buffer $(t_{1/2} = 7.2 \text{ h}).$

As a test of the ability of an acyl pyrophosphate to inactivate an enzyme that utilizes a related substrate, farnesyl synthetase from yeast (EC 2.5.1.1) was incubated with 0.25 mM isopentenoyl pyrophosphate.





This enzyme is likely to contain nucleophilic groups in its substrate binding site.¹⁵ Activity was monitored by using an acid lability assay with geranyl pyrophosphate and [1-14C]isopentenyl pyrophosphate.¹⁹ The enzyme lost all activity during a 2-min preincubation at 37 °C (the kinetics of the process were not determined due to the complexity of the assay). Activity could not be recovered by addition of excess substrate. Preincubation with large amounts of substrate protects the enzyme from inactivation. A schematic model for the inactivation reaction is shown in Figure 1.

Addition of the hydrolysis products of isopentenovl pyrophosphate, isopentenoic acid, and inorganic pyrophosphate under the same conditions gave no inactivation, although competitive inhibition (due to the pyrophosphate¹⁹) was observed. The specificity of inhibition is further demonstrated by our observation that acetyl pyrophosphate does not inactivate the enzyme (concentrations up to 5 mM).

These results indicate that acyl pyrophosphates can be conveniently prepared and possess reaction patterns that permit them to be used as enzyme inactivators. Detailed evaluations of these materials are necessary to determine their potential for specific applications.20

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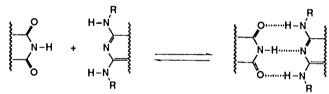
(20) Preliminary experiments (Huang, Z., unpublished) with isopentenyl pyrophosphate:dimethylallyl pyrophosphate isomerase indicate that isopentenoyl pyrophosphate is a reversible competitive inhibitor, consistent with a mechanism in which no nucleophile approaches the pyrophosphate ^{5,21} (21) Muelenbacher, M.; Poulter, C. D. J. Am. Chem. Soc. **1985**, 107, 8307.

New Molecular Complements to Imides. Complexation of Thymine Derivatives

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Molecular recognition of imides (Scheme I) is of interest in Scheme I



chromatographic resolutions,¹ thymine receptors,² liquid crystals,³ and molecular tapes and sheets,⁴ as well as theoretical⁵ and experimental⁶ evaluations of secondary effects in hydrogen bonding. We describe here new systems for base-pairing to thymines which offer unusual affinities and promise for the catalysis of reactions involving thymines.

The receptors are prepared from the readily available xanthene-1,8-dicarboxylic acid 1, and they resemble those derived from Kemp's triacid⁷ but have more spacious interiors.⁸ Esterification with phenol (DCC, CH_2Cl_2 , 0 °C) or naphthylethanol gave the mono esters⁹ 2 (Scheme II). Activation (SOCl₂) and then coupling with suitable amines gave the amide esters 3a-c (not shown), which were heated with biguanide (2 equiv) in ethanol to give the receptors 4a-c (30-40% overall from 1).9 The ester 4d was prepared from 2a by sequential treatment with SOCl₂ and

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(9) All new compounds were characterized by high-resolution NMR and MS and IR spectra. Mp (°C) (4a) 284-285, (4b) >310, (4c) 168-169, (4d) 238-2390, (4e) 283-285, (9) 163-164, (10) 222-223.

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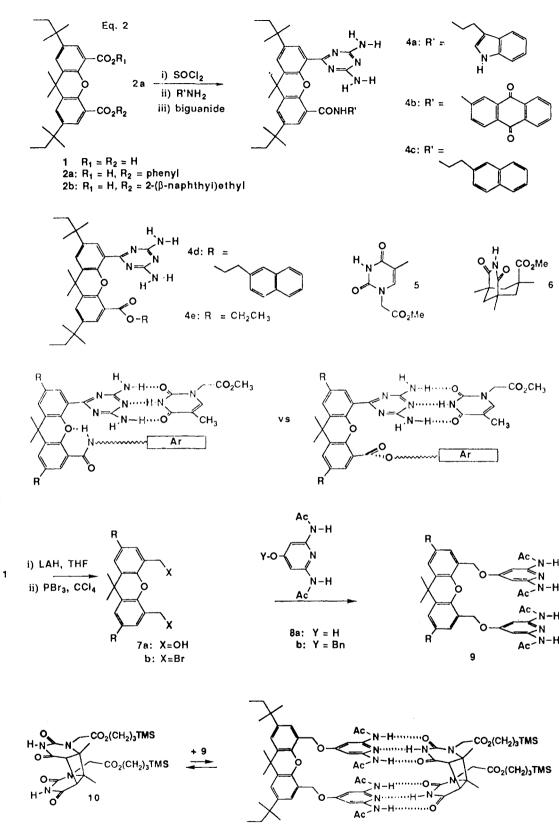
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Scheme III

Scheme IV

Scheme V



biguanide, while the ethyl ester triazine 4e was prepared by treatment of 1 with $SOCl_2$ and then biguanide in EtOH. The imides used were the thymine-derived methyl ester 5 and the saturated imide 6 obtained from Kemp's triacid.¹⁰

The affinities were determined by titrations in $CDCl_3$ using NMR; the results are given in Table I. Stacking can contribute up to an order of magnitude in binding affinities at room temperature. The amides are especially well suited for this as the intramolecular hydrogen bonds bring the side chains (bearing the aryl surfaces) close to the site of base-pairing (Scheme III).

Ditopic systems are also available. Reduction of 1 (LAH/THF) gave the diol 7a, from which the dibromide 7b was obtained by

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Table I. Association Constants

receptor	guest	K _a	receptor	guest	K
4a	5	2500	4d	5	1100
4b	5	6700	4e	5	670
4 c	5	2000	4 e	6	340

treatment with PBr3 in CCl4. Condensation with 2,6-bis(acylamino)-4-pyridone $8a^{2,11}$ (K₂CO₃, DMF) gave 9 (Scheme IV). Binding to 5 gave a 2:1 complex,¹² while the thymine photodimer 10¹³ gave a 1:1 complex ($K_a = 4800 \text{ M}^{-1}$). Model studies for the photolyase system¹⁴ and template synthesis of thymine derivatives are currently underway.

Acknowledgment. We are grateful to the National Institutes of Health for financial support of this research.

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11, 126. (12) The statistically corrected association constants for 9 with 5 were K_1 = 1250 and $K_2 = 400$; for 8b with 5, $K_4 = 960$. (13) The dicarboxylic acid (Cochran, A. G.; Sugasawara, R.; Schultz, P. G. J. Am. Chem. Soc. 1988, 110, 7888-7890) was esterified with (tri-methylsilyl)propanol using carbonyldiimidazole. (14) Hirst, S. C.; Hamilton, A. D. Tetrahedron Lett. 1990, 31, 2401-2404. Hamilton A. D. L. Chem. Commun. 1990, 207-200.

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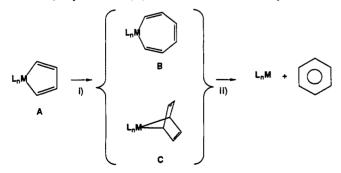
An η^4 -Benzene Species Mediates Acetylene Cyclotrimerization

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The mechanism of alkyne cyclotrimerization is widely discussed,¹⁻⁹ but most¹⁰ experiments do not strongly distinguish between the generally considered two paths (via B and C) from metallacyclopentadiene (A) to free arene. We now report results



wherein systematic variation of M from Rh to Ir, with $L_n =$ $MeC(CH_2PPh_2)_3$ (triphos), appears to eliminate intermediate B, yet C is a still an imperfect representation of the mechanism.

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We reported earlier¹¹ that a species isolated and spectroscopically characterized as the complex (triphos)RhCl(η^2 -C₄H₄) (Figure 1a)¹² is catalytically active for acetylene cyclotrimerization (6 turnovers/h at 1 atm of C_2H_2 and 25 °C). No additional intermediates were detected and, in particular, no experimental evidence was provided for distinguishing between paths B and C. However, when HCCH is passed through a THF solution of $(triphos)IrCl(C_2H_4)$ at 25 °C in the presence of a chloride scavenger¹³ (TIPF₆), solution ¹H and ³¹P^{[1}H] NMR data show the production of $(triphos)Ir(C_6H_6)^+$, which can be isolated as its BPh₄ salt. The structure¹⁴ of this cation (Figure 1b) shows it to be an 18-electron species with η^4 coordination of the C₆H₆ ligand. The dihedral angle within the bent benzene is 134.8°. The nonbonded carbons, C50 and C51, have Ir/C distances longer than 2.97 Å, and they are connected by a localized double bond of length 1,36 (3) Å. The observed C-C bond lengths share with all previous η^4 -benzene structures¹⁵ the C/C bond length pattern shown in D. This is symptomatic of a large degree of backbonding,^{16,17} which is appropriate for a 5d metal ligated by three electron-donating ligands (triphos).18



D

The variable-temperature ³¹P{¹H} NMR spectra in CDCl₃ show (triphos)Ir(η^4 -C₆H₆)⁺ to be fluxional. The rapid-exchange spectrum (>313 K) shows a single line, which transforms at low temperature to an AM₂ pattern. Simulation (DNMR3) yields $(253-313 \text{ K}) \Delta H^* = 10.9 \pm 0.3 \text{ kcal/mol and } \Delta S^* = -25 \pm 1$ cal K⁻¹ mol⁻¹. Variable-temperature ¹H NMR spectra in CDCl₃ show one broad resonance (5.18 ppm) for the C_6H_6 ring at 323

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(12) Crystal data for C₄₅H₄₃ClP₃Rh-CH₂Cl₂0.5C₇H₁₆ (-155 °C): a = 10.133 (1) Å, b = 23.620 (4) Å, c = 18.682 (3) Å, $\beta = 93.62$ (0)° with Z = 4 in space group $P2_1/c$. R(F) = 0.063 and $R_w(F) = 0.061$ for 2038 reflections with $F > 3\sigma(F)$ and anisotropic thermal parameters on Rh, Cl, and P.

(13) TIPF₆ is not required for cyclotrimerization by the more labile rhodium analogue.

(14) Crystal data for $C_{71}H_{65}BrIrP_{2}\cdot C_{5}H_{10}O$ (-155 °C): a = 16.471 (6) Å, b = 17.126 (6) Å, c = 12.030 (4) Å, $\alpha = 101.22$ (2)°, $\beta = 93.61$ (2)°, $\gamma = 75.46$ (1)°, with Z = 2 in space group PI. R(F) = 0.0845 and $R_w(F) = 0.0106$ (2)°, $\gamma = 10.0216$ (2)°, γ 0.0819 for 7211 absorption-corrected reflections with $F > 3\sigma(F)$ and anisotropic thermal parameters.

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(18) This pattern of C/C bond lengths is not seen in (triphos)M(butadi-ene)⁺ species of group VIII metals and thus is specific to the η^4 -benzene ligand in an electron-rich environment.

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